

The Effects of Time-Varying Temperature on Delays in Genetic Networks*

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Abstract. Delays in gene networks result from the sequential nature of protein assembly. However, it is unclear how models of gene networks that use delays should be modified when considering time-dependent changes in temperature. This is important, as delay is often used in models of genetic oscillators that can be entrained by periodic fluctuations in temperature. Here, we analytically derive the time dependence of delay distributions in response to time-varying temperature changes. We find that the resulting time-varying delay is nonlinearly dependent on parameters of the time-varying temperature such as amplitude and frequency; therefore, applying an Arrhenius scaling may result in erroneous conclusions. We use these results to examine a model of a synthetic gene oscillator with temperature compensation. We show that temperature entrainment follows from the same mechanism that results in temperature compensation. Under a common Arrhenius scaling alone, the frequency of the oscillator is sensitive to changes in the mean temperature but robust to changes in the frequency of a periodically time-varying temperature. When a mechanism for temperature compensation is included in the model, however, we show that the oscillator is entrained by periodically varying temperature even when maintaining insensitivity to the mean temperature.

Key words. systems biology, genetic networks, time-varying delay

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1. Introduction. Biochemical reaction rates, like all chemical reaction rates, are sensitive to changes in temperature, and this effect is captured mathematically by the Arrhenius equation [2, 26]. Temperature-dependent rates can also alter the dynamics of gene regulatory networks. Previous studies have examined how gene networks behave at various temperatures, and, in general, their dynamics speed up with increasing temperature [34]. For example, the cell doubling time in root meristems of *Zea mays* decreases 21-fold from a 3–25°C increase in temperature [10]. In nature, variations in temperature often occur at time scales that are much slower than typical time scales of gene networks. However, time-varying temperatures have been shown to impact gene networks, particularly in the temperature entrainment of circadian oscillators. In such systems, the diurnal variations in temperature happen at the

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same time scale as the circadian network. For instance, circadian oscillators can be entrained by time-varying temperatures that cycle within a period close to 24 hours [5, 24, 33, 44].

In models of gene networks, dynamical delay has been used to model the sequential assembly of messenger RNA (mRNA) and then protein. Nucleic acids must be added one by one to the growing mRNA chain, while amino acids are joined end to end with peptide bonds to create a protein. In each case, the large chain of linear reactions can be compactly modeled either with a discrete delay term or as a distributed delay term [4, 16]. The incorporation of delay greatly simplifies models of genetic oscillators while simultaneously maintaining qualitative similarities to experimental data [7, 19, 38, 39]. Delay-based models play a central role in understanding the origin of oscillations in genetic networks [28, 29] and other nonlinear systems [16, 20]. For constant temperatures, the delay time or distribution can be scaled with the Arrhenius equation, just as the reaction rates. However, less is known about how time-varying temperatures influence delays in such analyses.

Here, we investigate how time-varying temperatures affect delays in genetic networks. We first derive how time-dependent temperature affects the delay term. To do this, we assume that delays arise from a sequence of first-order reactions that can be modeled as an aggregate delay. Although delay in gene networks is the result of the sequential assembly of first mRNA [6] and then protein [30] (see Figure 1), we lump these delays into one term and represent protein production delay as a reduction of a linear chain of reactions. Each reaction in the sequence is then scaled by a common time-dependent Arrhenius factor. Since changes in temperature influence each biochemical step in the sequence that constitutes the delay, the value of the delay time will change. From these assumptions, we derive an expression for the time-dependent distribution of delay times. We analyze changes in phase shift and amplitude of the resulting time-varying delay as a function of parameters of a sinusoidally time-varying rate-coefficient induced by temperature changes. We find a nonlinear relationship and, furthermore, find specific cases for which a delay can remain approximately time-invariant under time-varying conditions.

The effects of temperature on oscillators becomes important in the study of circadian clocks and is typically inferred through analysis of system response to single step changes in temperature or a single cycle [24]. We incorporate our findings into a model of a synthetic gene oscillator with temperature compensation presented by Hussain et al. [19]. In that paper, Hussain et al. presented experimental and computational results of a synthetic gene oscillator that has the same period regardless of the temperature. However, they only considered constant temperatures. Here, we extend their computational model to include time-varying temperatures. We find that, when the temperature varies sinusoidally in time, the oscillator can be entrained by temperature, but that this entrainment does not occur in the absence of the temperature compensation mechanism. In other words, we find that the temperature compensated oscillator is insensitive to changes in the mean temperature but also entrained by periodically varying temperatures, a property explained and observed in circadian oscillators.

2. Characterization of time-varying delays. We begin by approximating protein production with a linear sequence of reactions (see Figure 1). Transcription can be modeled as a sequence of independent one-nucleotide reaction steps as has been done by Arkin, Ross, and McAdams in [1]. Furthermore, Bel, Minsky, and Nemenman [4] investigated more general

cases by including degradation due to cellular division and considering reversible reactions. They found that in the limiting case of decreasing relative variance all systems with a forward bias behaved similarly. In this paper we consider only a simplified model, i.e., one with irreversible reactions and zero probability of partial transcripts. Since cellular division times are much longer than transcriptional delays, we also neglect dilution due to cell division, but this could be incorporated through a correction factor on the overall production rate as explained by O'Brien, Italic, and Bennett in [31]. In this case, we consider the corresponding generalized mass action model, the dynamics of which can be modeled by

$$(2.1) \quad \dot{x}_j = -a(t)(x_j(t) - x_{j-1}(t)) \quad \text{for } j = 1, \dots, N,$$

where $x_i(t)$ is the concentration of the i th species at time t , $x_0(t)$ is the time-varying concentration of the initial complex, $a(t)$ is the time-varying rate coefficient, and the overdot represents differentiation with respect to time. The effects of time-varying temperatures can be reflected in the time-varying rate coefficients. From this we deduce the effects of time-varying temperatures on the delay distribution, i.e., the time it takes to go from the initial complex, x_0 , to mature protein, x_N .

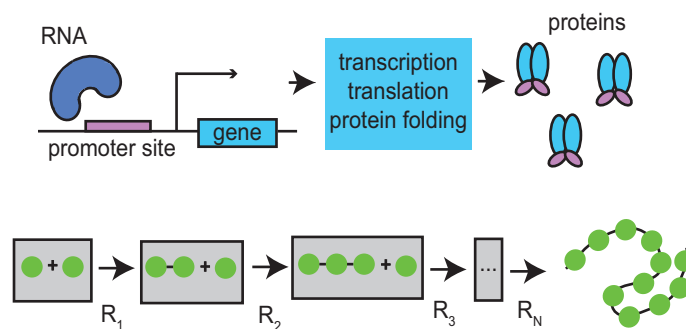


Figure 1. Modeling of delays in protein production. *Transcriptional delays (top) are modeled by a sequence of chemical reactions (bottom) with common reaction rate $a(t)$ for each reaction R_1, \dots, R_N .*

To find the distribution function, we first rewrite system (2.1) as

$$(2.2) \quad \dot{\mathbf{x}}(t) = \mathbf{A}(t) \mathbf{x}(t) + \mathbf{u}(t),$$

where $\mathbf{x}(t) = [x_N, x_{N-1}, \dots, x_1]^T$, $\mathbf{u}(t) = [0, \dots, 0, a(t)x_0(t)]$, and $\mathbf{A}(t) = a(t) \mathbf{J}_{-1,N}$. Here $\mathbf{J}_{-1,N}$ is the N -dimensional Jordan matrix with eigenvalues -1 ,

$$(2.3) \quad \mathbf{J}_{-1,N} = \begin{bmatrix} -1 & 1 & 0 & \dots & 0 \\ 0 & -1 & 1 & & \\ \vdots & & \ddots & \ddots & \\ & & & -1 & 1 \\ 0 & \dots & & 0 & -1 \end{bmatrix}.$$

Because $\mathbf{A}(t_1)$ commutes with $\mathbf{A}(t_2)$ for all (t_1, t_2) , we can write the general solution to (2.2) as

$$(2.4) \quad x(t) = \phi(t, t_0) x(t_0) + \int_{t_0}^t \phi(t, \sigma) u(\sigma) d\sigma,$$

where

$$(2.5) \quad \phi(t, t') = \exp \left(\int_{t'}^t \mathbf{A}(s) ds \right).$$

Without loss of generality we set $t_0 = 0$ and substitute $\sigma = t - \tau$. If we assume $x_j(t_0) = 0$ for $j = 1, \dots, N$, the solution reduces to

$$(2.6) \quad x(t) = \int_0^t e^{\alpha(t, \tau) \mathbf{J}_{-1, N}} u(t - \tau) d\tau,$$

where $\alpha(t, \tau) \doteq \int_{t-\tau}^t a(s) ds$. The exponential can be computed and is given by

$$(2.7) \quad e^{\alpha(t, \tau) \mathbf{J}_{-1, N}} = \begin{bmatrix} e^{-\alpha(t, \tau)} & \alpha(t, \tau) e^{-\alpha(t, \tau)} & \dots & \frac{\alpha(t, \tau)^{N-2}}{(N-2)!} e^{-\alpha(t, \tau)} & \vdots & \frac{\alpha(t, \tau)^{N-1}}{(N-1)!} e^{-\alpha(t, \tau)} \\ \hline & & & & \vdots & \frac{\alpha(t, \tau)^{N-1}}{(N-1)!} e^{-\alpha(t, \tau)} \\ & & \star & & \vdots & \\ & & & & \alpha(t, \tau) e^{-\alpha(t, \tau)} & \\ & & & & e^{-\alpha(t, \tau)} & \\ & & & & & \vdots \end{bmatrix},$$

where \star denotes nonzero entries that are irrelevant due to the structure of $u(t)$ and our desired output. In order to extract the expression relating the input $x_0(t)$ to the measured output $x_N(t)$, we multiply (2.6) by $C = [1, 0, \dots, 0]$ on the left-hand side and substitute the expression for $u(t)$, which gives the result

$$(2.8) \quad x_N(t) = \int_0^t h(t, \tau) x_0(t - \tau) d\tau,$$

where the function

$$(2.9) \quad h(t, \tau) = a(t - \tau) \frac{\alpha(t, \tau)^{N-1}}{(N-1)!} e^{-\alpha(t, \tau)}$$

is the impulse response function relating the output to the input of the system. We refer to the impulse function as the delay distribution corresponding to protein synthesis times. Although we are working with a deterministic system, we refer to the function as a delay distribution since it emerges from a stochastic process. Recall that the systems of ODEs from which the expression was derived is the generalized mass action model of a sequence of stochastic reactions. The impulse function describes the average dynamics of that process. The delay distribution must have the constraint $\int_0^\infty h(t, \tau) d\tau = 1$ for any time t . This constraint is always satisfied by the physics of the problem. The integral can be shown to equal one when

$a(t - \tau) > \epsilon$ for some $\epsilon > 0$, that is, when the reaction rate at all times is positive definite. To show this we express the integral as a line integral:

$$\begin{aligned}
 \int_0^\infty h(t, \tau) d\tau &= \int_0^\infty a(t - \tau) \frac{\alpha(t, \tau)^N}{(N)!} e^{-\alpha(t, \tau)} d\tau \\
 (2.10) \qquad &= \int_0^\infty \frac{\alpha(t, \tau)^N}{(N)!} e^{-\alpha(t, \tau)} \left| \frac{\partial}{\partial \tau} (\alpha(t, \tau)) \right| d\tau \\
 &= \int_C \frac{s^N}{N!} e^{-s} ds,
 \end{aligned}$$

where the curve C is the domain of integration that is defined by $\alpha(t, \tau)$ for t held fixed. Note that the expression in the last line is an integral over the Erlang distribution, which is equal to one when integrated along the curve $C \equiv a\tau$. For the line integral in (2.10) to equal one, $\alpha(t, \tau)$ must be an injective function in τ , meaning $d\alpha(t, \tau)/d\tau = a(t - \tau) > 0$ (which implies $a(t) > 0$ for all t) with $\alpha(t, 0) = 0$ and $\lim_{\tau \rightarrow \infty} \alpha(t, \tau) = \infty$. By the definition of $\alpha(t, \tau)$ ($\alpha(t, \tau) \doteq \int_{t-\tau}^t a(s) ds$), we see that a positive definite $a(t)$ in turn satisfies the latter conditions.

For the purpose of demonstration we consider a sinusoidally time-varying rate coefficient $a(t) = \delta_p a_0 \sin(\omega t) + a_0$ with $0 < \delta_p < 1$, assuming the dynamics are induced from an appropriate time-varying temperature. Next, we show conditions under which this is a good approximation for a sinusoidally time-varying temperature. When $a(t) \equiv \text{const}$ (i.e., $a(t) = a_0$), (2.9) is the *Erlang* distribution [27] with mean N/a . If $a(t)$ is not constant, the delay distribution will be a function of time. We define the variable E to be $E = N/a_0$ for the time-varying case and refer to E as the expected delay. Figure 2(a)–(b) shows the delay distribution $h(t, \tau)$ for different values of N (holding $E = N/a_0$ constant), comparing the time-invariant case to the time-varying case. Note that, unlike the time-invariant case (Figure 2(a)), the distribution in the time-varying case (Figure 2(b)) need not be unimodal, especially for small N . For a large N , the distribution function drops off quickly away from the mean and damps out any periodic fluctuations induced by the factor $a(t - \tau)$ in (2.9). However, for small N , we see the effects of the periodically varying reaction rate in the shape of the distribution. This multimodal distribution of protein synthesis times for transcription initiation at a given time t emerges from the underlying stochastic process combined with sensitivity to timing. For example, there may be a set of transcripts which happen to complete right before temperatures plunge again, leaving behind another set of transcripts to be significantly delayed. Figure 2(c) shows how the time-varying distribution changes with time for fixed N and a time-varying rate coefficient $a(t)$. Note that the distribution becomes unimodal as N increases.

Next, consider the limit as the number of reactions within the sequence tends to infinity. In the time-invariant case, one would consider the limit as $N \rightarrow \infty$ such that the mean of the distribution N/a remains constant. Taking this limit reduces the distribution function to the Dirac delta function $\delta(\tau - N/a)$, which has been shown in other work [4, 11]. To investigate the time-varying case, we assume that $a(t) = a_0 f(t)$, where $a_0 > 0$ and $f(t)$ is a positive definite, bounded function of time, which agrees with the sinusoidally varying function $a(t)$ above. In this case, if we take the limit $N \rightarrow \infty$ with the constraint $E = N/a_0 \equiv \text{const}$, the ratio $N/\alpha(t, \tau)$ remains finite for finite τ .

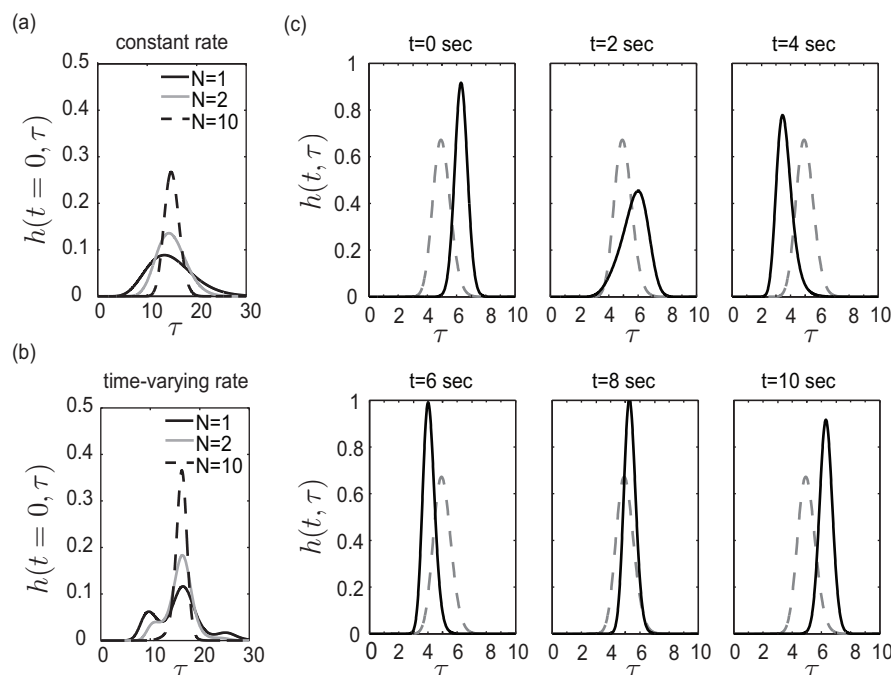


Figure 2. Delay distribution for different values of N with $E = 15$ for a constant and time-varying rate a . (a) Delay distribution for different values of N with $E = 15$ for a constant rate coefficient a . (b) Delay distribution for different values of N with $E = 15$ for a time-varying rate coefficient $a(t) = .5 a_0 \sin(\omega t) + a_0$ at time $t = 0$. (c) Distribution as a function of time with $a(t) = a_0 \delta_p \sin(\omega t) + a_0$, $N = 100$, $(N + 1)/a_0 = 15$, $\delta_p = .5$, and $\omega = 2\pi/20$. The dashed line indicates the nominal time-invariant distribution with $a(t) = a_0$.

In summary, we find that there exists a unique delay τ_{eff} such that

$$(2.11) \quad \lim_{N \rightarrow \infty} h(t, \tau) = \begin{cases} 0, & \tau \neq \tau_{\text{eff}}, \\ \infty, & \tau = \tau_{\text{eff}}. \end{cases}$$

The derivation of these results can be found in Appendix A. Therefore, with the integral over the function equal to one, in the limit as $N \rightarrow \infty$ such that $N/a_0 = E_0$ the distribution is approximated by a delta function centered at τ_{eff} (i.e., $\lim_{N \rightarrow \infty} h(t, \tau) \approx \delta(t - \tau_{\text{eff}}(t))$), which is necessarily a function of $a(t)$, and therefore time-varying. From the derivation we find that this unique delay $\tau_{\text{eff}}(t)$ can be found by solving

$$(2.12) \quad \int_{t-\tau_{\text{eff}}}^t f(s) ds = E$$

for τ_{eff} . Note that $\tau_{\text{eff}}(t)$ can be computed with only the expected delay $E = N/a_0$ and the time-varying function $f(t)$. Also, since $f(s)$ is positive definite, we can guarantee a single solution τ_{eff} for every time t . The effective delay τ_{eff} is computed such that the integral remains constant at E . Note that the area under the curve is zero for $\tau_{\text{eff}} = 0$ and monotonically increases with increasing τ_{eff} .

We now apply the method to investigate delays under periodically time-varying temperatures. For a time-varying temperature

$$(2.13) \quad T = \Delta T \sin(\omega t) + T_0$$

we can rewrite the Arrhenius equation $A(T) = A_0 e^{-\theta/T}$ in the form

$$(2.14) \quad A(\epsilon) = A_0 \exp\left(\frac{K}{1 + \epsilon \sin(\omega t)}\right),$$

where $\epsilon = \frac{\Delta T}{T_0}$ is nonnegative and $K = -\theta/T_0$. We assume $\epsilon \ll 1$ (i.e., $\Delta T \ll T_0$), and in the Taylor expansion we have

$$(2.15) \quad \begin{aligned} A(\epsilon) &\cong A(0) + \frac{A'(0)}{1!} \epsilon + \cdots \\ &= A(0) - K \sin(\omega t) \epsilon + \cdots \\ &= 1 + \frac{\theta}{T_0} \epsilon \sin(\omega t) + \cdots \end{aligned}$$

Here, A_0 is chosen such that $A(0) = 1$, i.e., $A_0 = \exp(\theta/T_0)$. Therefore, for small ϵ we can approximate the time-varying Arrhenius equation by

$$(2.16) \quad A(\epsilon) \approx 1 + \delta_p \sin(\omega t),$$

where $\delta_p = \frac{\theta}{T_0} \epsilon$. The time-varying rate coefficient for the reaction rates implicit in the delay is given by $a(t) = a_0 \cdot A(t)$.

For a sinusoidally varying rate coefficient $a(t) = a_0 \delta_p \sin(\omega t) + a_0$, using (2.12), the effective delay reduces to solving

$$(2.17) \quad \int_{t-\tau_{\text{eff}}}^t [\delta_p \sin(\omega s + \phi) + 1] ds = E,$$

where E is the expected delay. In the limit analyzed, changes in the delay are determined only from the expected delay without perturbation and the perturbation on the reaction rates. Also note that in the extreme limits of the frequency, we have

$$(2.18) \quad \tau_{\text{eff}} \approx \begin{cases} E & \text{for } \omega \rightarrow \infty, \\ \frac{E}{\delta_p \sin \phi + 1} & \text{for } \omega \rightarrow 0. \end{cases}$$

Taking the integral in (2.17), the solution can be shown to solve the system

$$(2.19) \quad \tau = E + \frac{\delta_p}{\omega} \cos(\omega t + \phi) - \frac{\delta_p}{\omega} \cos(\omega t + \phi - \omega \tau).$$

In this case the effective delay at each time t must be solved numerically. Figure 3 shows the delay as a function of time for a periodically varying temperature. The solution is found numerically for a discretized range of time. In Figure 3 we consider the expected delay $E = 13.5$ min.

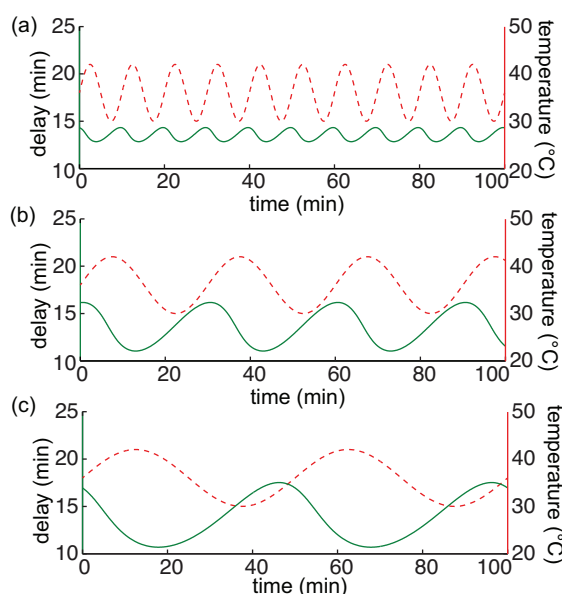


Figure 3. Amplitude of the delay changes with the period of the time-varying temperature. *Time-varying delays (green solid lines) corresponding to various time-varying temperatures $T(t) = \Delta T \sin(\omega t) + T_0$ (red dashed lines).* Parameter values here are $\theta = 4500K$, $E = 13.5 \text{ min.}$, $\Delta T = 6^\circ C$, $T_0 = 36^\circ C$, and $\delta_p = .28$. (a) $\omega = \frac{2\pi}{10}$. (b) $\omega = \frac{2\pi}{30}$. (c) $\omega = \frac{2\pi}{50}$.

We now consider the effects of changing parameters δ_p and ω on the time-varying delay. Figure 4 shows analysis of the calculated time-varying delay as a function of various parameters. Figure 4(a) shows an example of the time-varying function $\tau_{\text{eff}}(t)$. In Figure 4(b)–(d), we look at how the mean and amplitude of the time-varying function τ_{eff} changes as we change the expected delay E , the relative perturbation δ_p , and the frequency ω of the time-varying rate function $a(t)$. Most of the results are in line with intuition. For example, the amplitude of $\tau_{\text{eff}}(t)$ increases with an increase in the relative perturbation δ_p and decreases as the frequency ω of $a(t)$ increases. If the environmental conditions change too quickly, the system does not effectively respond. An unexpected result is the nonmonotonic behavior of the function $\tau_{\text{eff}}(t)$ as the mean delay changes. Figure 4(b) implies that τ_{eff} remains constant when the mean delay is exactly equal to the period of $a(t)$. With this observation we note that if $\omega = 2\pi n/T$ for any positive integer n in (2.19), then we always have the solution $\tau_{\text{eff}}(t) = T$. This suggests that delays are minimally affected by sinusoidally time-varying reaction rates when the mean delay is an integer multiple of the period. We see the result of this in Figure 4(c) as well.

It is apparent from Figure 4 that the dependence of τ_{eff} on the sinusoidally time-varying temperature can be nonlinear. In this respect, we analyze the phase shift between the sinusoidally time-varying reaction rate $a(t)$ and the resulting time-varying delay $\tau_{\text{eff}}(t)$. In Figure 5(a)–(c), we look at how the phase shift changes as we change the expected delay E , the relative perturbation δ_p , and the period of the time-varying rate function $a(t)$. Before calculating phase shift, we account for the fact that the reaction rate and delay functions are initially 180° out of phase because the delay decreases when the reaction rate $a(t)$ increases. Also, since $\tau_{\text{eff}}(t)$ is not a perfect sinusoid, we calculate phase shift based on the distance

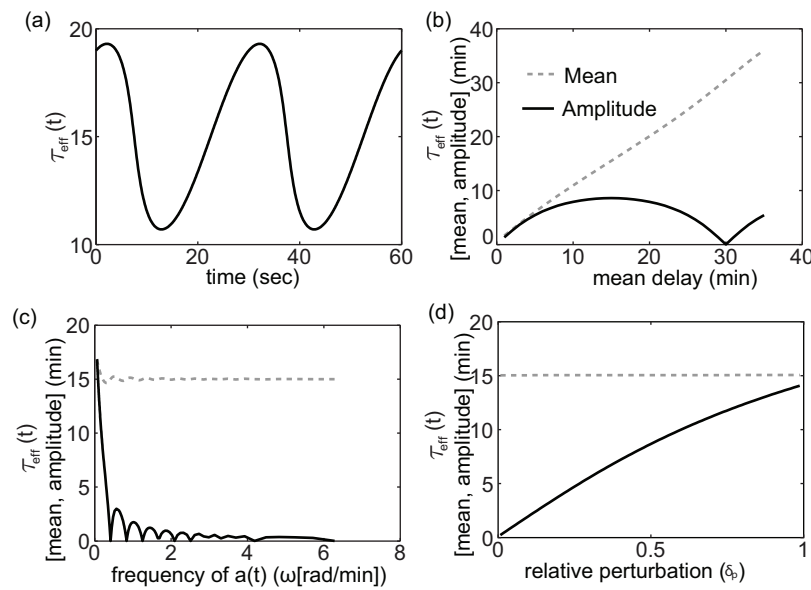


Figure 4. Mean and peak-to-peak amplitude of τ_{eff} as a function of δ_p , E , and ω . (a) τ_{eff} as a function of time with $\omega = \pi/15$, $\phi = 0$, $E = 15$, and $\delta_p = .5$. (b) τ_{eff} as a function of the expected delay with $\omega = \pi/15$, $\phi = 0$, and $\delta_p = .5$. (c) τ_{eff} as a function of the frequency of $a(t)$ with $\phi = 0$, $E = 15$, and $\delta_p = .5$. (d) τ_{eff} as a function of the relative perturbation δ_p with $\omega = \pi/E$, $\phi = 0$, and $E = 15$.

between peaks. In general we find an increase in phase shift with an increase in expected delay E , the relative perturbation δ_p , and the period of the time-varying rate function $a(t)$. However, Figure 5(b)–(c) shows an existence of discontinuities in the phase shift. There is a 180° phase jump when the expected delay equals the period. Recall that the amplitude of $\tau_{eff}(t)$ becomes zero when the delay is an integer multiple of the period. As the response $\tau_{eff}(t)$ crosses this critical point, there is a 180° phase shift as the amplitude of the response becomes nonzero again. Furthermore, the frequency of discontinuities increases on the log scale as a function of frequency and expected delay. We show trends in a limited range of frequencies and expected delays in Figure 5(b)–(c) to help highlight where the nonlinearities come from and demonstrate the nontrivial behavior.

3. Temperature entrainment of a dual-feedback oscillator. We now consider the entrainment properties of a temperature compensated dual-feedback oscillator presented by Hussain et al. [19]. The oscillator, as depicted in Figure 6, can be modeled as [19]

$$\begin{aligned}
 (3.1) \quad \frac{1}{A(T)} \cdot \frac{dx}{dt} &= \frac{\alpha_x \left(\eta + \frac{y(t-\tau_y)}{C_y} \right)}{\left(1 + \frac{y(t-\tau_y)}{C_y} \right) \left(1 + \frac{x(t-\tau_x)}{C_x(T)} \right)^N} - \beta x(t) - \frac{\gamma_x x(t)}{R_0 + x(t) + y(t)}, \\
 \frac{1}{A(T)} \cdot \frac{dy}{dt} &= \frac{\alpha_y \left(\eta + \frac{y(t-\tau_y)}{C_y} \right)}{\left(1 + \frac{y(t-\tau_y)}{C_y} \right) \left(1 + \frac{x(t-\tau_x)}{C_x(T)} \right)^N} - \beta y(t) - \frac{\gamma_y y(t)}{R_0 + x(t) + y(t)},
 \end{aligned}$$

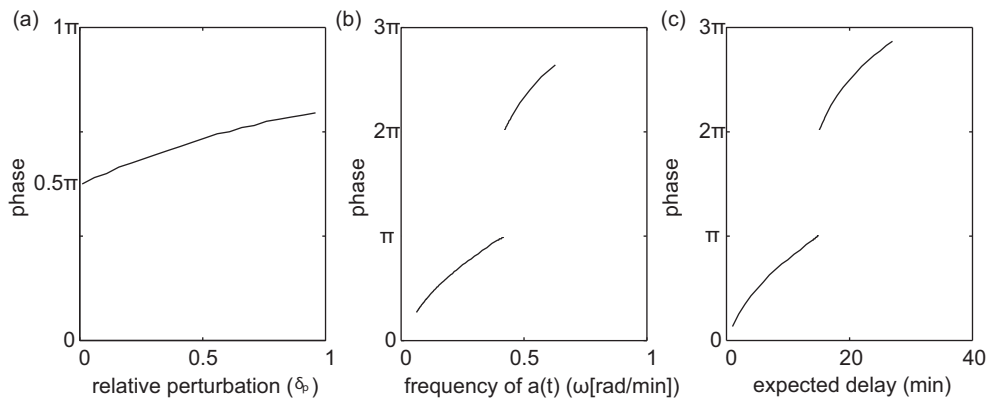


Figure 5. Phase shift between $a(t)$ and $\tau_{\text{eff}}(t)$ as a function of δ_p , E , and ω . (a) Phase shift as a function of δ_p with $E = 15$ and $\omega = 2\pi/30$. (b) Phase shift as a function of ω with $E = 15$ and $\delta_p = .5$. (c) Phase shift as a function of E with $\omega = 2\pi/50$ and $\delta_p = .5$.

where x and y are the concentrations of the repressor (LacI) and the activator (AraC); α_x and α_y are the maximal transcription initiation rates for x and y , respectively; C_x and C_y are the binding affinities of LacI and AraC to the promoter, respectively; β is the dilution rate due to cellular growth; η is a measure of the strength of the positive feedback loop; R_0 , γ_x , and γ_y are Michaelis–Menten constants for enzymatic decay of the proteins; τ_x and τ_y are temperature-dependent delay times for the production of LacI and AraC, respectively; and $A(T)$ is the common Arrhenius scaling of all reaction rates. Additionally, the Arrhenius scaling term has the form $A(T) = A_0 e^{-\theta/T}$, where θ is the temperature scale. Note that increasing temperature increases the scaling coefficient $A(T)$ and hence speeds up the dynamics of the system. In Hussain et al. [19], the authors scale the delay by the Arrhenius constant when predicting dynamics at varying temperatures (the temperatures are held constant for each assay). In this case, we consider predictions under time-varying temperatures defined by (2.13). We use the method derived in section 2 to determine the resulting time-varying delay. The binding affinity of LacI, $C_x(T)$ is also a function of the temperature

$$(3.2) \quad C_x(T) = (C_{x,\max} - C_{x,\min}) \frac{(T/T_{\text{lac}})^b}{1 + (T/T_{\text{lac}})^b} + C_{x,\min}$$

and provides the mechanism for temperature compensation in the oscillator [19]. $C_{x,\min}$ and $C_{x,\max}$ are the minimum and maximum binding affinities of LacI to its promoter. T_{lac} is the temperature at which $C_x(T)$ is half-maximal and b is a Hill coefficient.

In Hussain et al. [19], the period of the genetic oscillator is shown experimentally to remain largely unaffected by changes in constant temperature due to a temperature sensitive LacI mutant, which is modeled by the temperature-dependent binding affinity $C_x(T)$. We now consider entrainment properties of the circuit with time-varying temperatures in silico. In order to compare entrainment properties to a system without such a temperature-dependent mechanism, we consider the temperature-invariant binding affinity

$$(3.3) \quad C_x(T) = C_x(T_0)$$

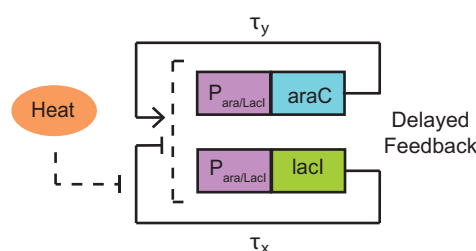


Figure 6. Schematic of the temperature compensating oscillator [19].

and compare the two models. First, we study system (3.1), (3.2) and compare results with system (3.1), (3.3). In this case temperature effects are introduced solely through an Arrhenius scaling and implicitly through the time-varying delay. We drive the system with a time-varying temperature described by (2.13) with $\Delta T = 2^\circ\text{C}$. From this we can approximate the time-varying Arrhenius scaling using (2.16) with $\theta = 4500\text{K}$. Then the delay is calculated at each time step by solving (2.19). Details of the simulations are found in Appendix B. In Figure 7(a) we fix the frequency of the temperature at $\omega = \frac{2\pi}{50}$ rad/min and vary the mean temperature T_0 in the two models in order to verify the temperature compensating property achieved through the temperature-dependent LacI mutant. Without the temperature compensating mechanism the frequency of oscillations changes linearly with the mean temperature but remains constant with the LacI mutant. In Figure 7(b) we fix the mean temperature at $T_0 = 36^\circ\text{C}$ and vary the frequency ω to study frequency entrainment for the same system with and without the temperature compensating mechanism. It is clear that the system entrains only under the influence of the temperature sensitive promoter. A common Arrhenius scaling alone does not allow for frequency or phase entrainment. The same mechanism that provides temperature compensation (insensitivity to changes in mean temperature) also makes the system sensitive to temperature dynamics, achieving entrainment. This is in agreement with circadian clocks as well [5].

Finally, we consider a stochastic analogue of system (3.1), (3.2) with a distributed delay as a more realistic model. Discrete delays are used as a simplifying approximation but do not occur in nature. We implement a sequence of reaction equations to model the delay (the same reaction equations used to arrive at (2.1)). Therefore, the probability density function describing the time required to complete transcription should be described by (2.9). The stochastic model is simulated using the standard Gillespie algorithm [12]. The details of the model can be found in Appendix C. The results are shown in Figure 8, where we look at response of the system to varying mean temperatures and frequencies as done above. Each point represents the average steady state period across 20 simulations with error bars representing a standard deviation above and below the mean. We see a response similar to that seen in the deterministic model.

4. Conclusion. It was found that periodic temperature fluctuations induce periodically time-varying delays. The effects of a time-varying temperature on delays within a genetic network can be highly nonlinear, and so the delay cannot be simply scaled by an Arrhenius coefficient in this case. With this, we investigated properties of a delay-based model of a

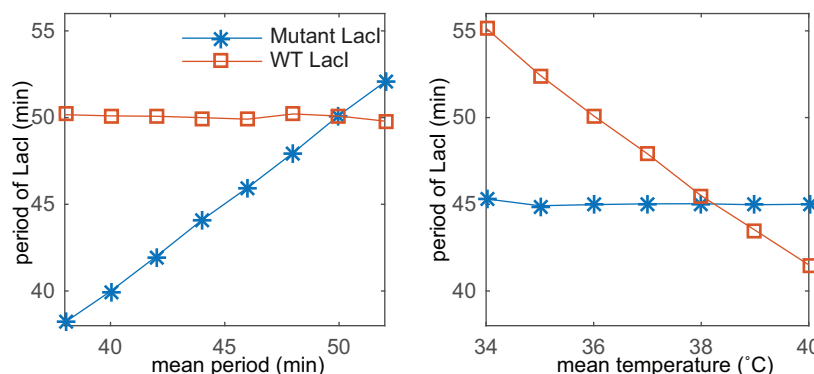


Figure 7. Entrainment of the synthetic gene oscillator with (Mutant LacI) and without a temperature sensitive promoter (WT LacI). (a) *Frequency entrainment of the circuit with and without temperature compensation for $\Delta T = 2^\circ\text{C}$ and $T_0 = 36^\circ\text{C}$.* (b) *Period of the circuit with and without temperature compensation for $\omega = 2\pi/50\text{ min}^{-1}$, $\Delta T = 2^\circ\text{C}$, and different mean temperatures T_0 in (2.13).*

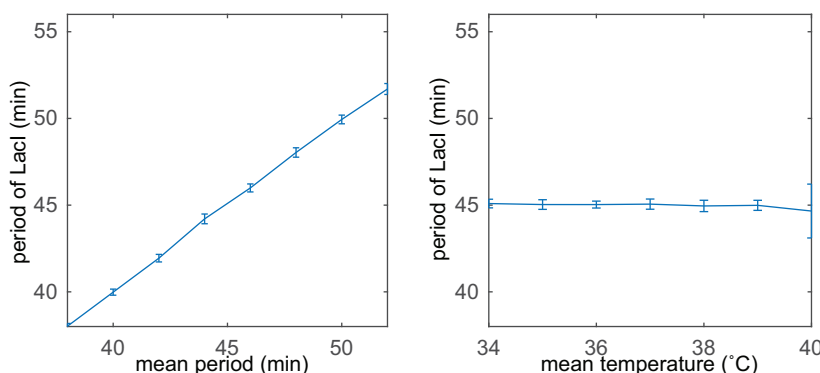


Figure 8. Gillespie simulation of the synthetic gene oscillator with a temperature sensitive promoter showing entrainment. (a) *Frequency entrainment of the circuit with $\Delta T = 5^\circ\text{C}$ and $T_0 = 36^\circ\text{C}$.* (b) *Period of the circuit for $\omega = 2\pi/50\text{ min}^{-1}$, $\Delta T = 5^\circ\text{C}$, and different mean temperatures T_0 in (2.13).*

temperature sensitive oscillator. This oscillator has been shown to exhibit temperature compensation; that is, the frequency of oscillation is insensitive to temperature variations. This was shown by analyzing the dynamics at different constant temperatures. Using the method derived, we were able to simulate the system under a periodically time-varying temperature. Simulations showed improved temperature compensating properties under the dynamically varying temperatures, over constant temperatures. Simulations also predicted reliable temperature entrainment. The frequency of protein expression coincided with that of the time-varying temperature.

We focused on properties important in circadian oscillators, namely, temperature com-

compensation and temperature entrainment. Ideally, a circadian oscillator should demonstrate properties of entrainment with insensitivity to changes in mean temperatures [9]. Here we highlight a case where the entrainment is a byproduct of the same mechanism which makes the system insensitive to changes in mean temperature. This is in agreement with Bodenstein, Heiland, and Schuster [5], where temperature entrainment was shown to naturally follow from circadian clock models tuned for temperature compensation through the Arrhenius coefficients. In the oscillator of Hussain et al. [19], there is an inherent tradeoff between robustness to unwanted temperature fluctuations about a mean and robustness to changes in mean temperatures, with the latter admitting temperature entrainment. Here, an understanding of the effects of temperature on delays eased the analysis of a delay-based model of a circuit with circadian clock-like properties.

Future work includes investigation of circadian oscillators, which have an intricate relationship with temperature. For instance, circadian oscillators exhibit temperature compensation [3, 35]; i.e., their periods do not vary with changes in the average temperature. Theorists have investigated methods of temperature compensation in models of circadian oscillators, often minimizing the effects of Arrhenius-scaled rate constants [9, 15, 17, 18, 37, 40]. Periodic changes in temperature have also been implicated in the entrainment of circadian oscillators to the day/night cycle [24, 33, 36, 41, 44]. However, entrainment of circadian oscillators is most commonly associated with periodic changes in light, and mathematical models have been developed explaining this phenomenon [8, 13, 14, 21, 22, 23, 25, 32, 42, 43]. Less is understood about the role of temperature.

Appendix A. Time-varying distribution limit. Here, we provide details on taking the limit $N \rightarrow \infty$ with the constraint $N/a_0 = E$ on the time-varying distribution

$$(A.1) \quad h(t, \tau) = a(t - \tau) \frac{\alpha(t, \tau)^{N-1}}{(N-1)!} e^{-\alpha(t, \tau)}.$$

We will show that in the limit the distribution, while maintaining an integral equal to one (as shown in the main text), becomes zero everywhere and infinity at a single point. In summary, we find that there exists a unique delay τ_{eff} such that

$$(A.2) \quad \lim_{N \rightarrow \infty} h(t, \tau) = \begin{cases} 0, & \tau \neq \tau_{\text{eff}}, \\ \infty, & \tau = \tau_{\text{eff}}. \end{cases}$$

Applying Stirling's formula for large N , namely

$$(A.3) \quad N! \approx \sqrt{2\pi N} \left(\frac{N}{e}\right)^N,$$

to the distribution (A.1) gives

$$(A.4) \quad h(t, \tau) = a(t - \tau) \frac{\alpha(t, \tau)^N}{N!} e^{-\alpha(t, \tau)}$$

$$(A.5) \quad \approx \frac{a(t - \tau)}{\sqrt{2\pi N}} \left(\frac{e \alpha(t, \tau)}{N}\right)^N e^{-\alpha(t, \tau)},$$

which asymptotically converges to (A.1) in the limit as $N \rightarrow \infty$. Rearranging terms in (A.5) and making use of the substitution $a(t) = a_0 f(t)$ gives

$$\begin{aligned}
 h(t, \tau) &\approx \frac{a(t-\tau)}{\sqrt{2\pi N}} \left(\frac{\alpha(t, \tau)}{N} e^{1-\alpha(t, \tau)/N} \right)^N \\
 &= \frac{a_0 f(t-\tau)}{\sqrt{2\pi N}} \left(\frac{\alpha(t, \tau)}{N} e^{1-\alpha(t, \tau)/N} \right)^N \\
 &= \left(\frac{N}{E_0} \right) \frac{f(t-\tau)}{\sqrt{2\pi N}} \left(\frac{\alpha(t, \tau)}{N} e^{1-\alpha(t, \tau)/N} \right)^N \\
 &= \frac{f(t-\tau)}{E_0 \sqrt{2\pi}} \left(\frac{\alpha(t, \tau)}{N} e^{1-\alpha(t, \tau)/N} \right)^N N^{\frac{1}{2}}.
 \end{aligned}
 \tag{A.6}$$

We define

$$K(\tau) \doteq \frac{\alpha(\tau)}{N} e^{1-\alpha(\tau)/N} \tag{A.7}$$

and investigate the limit of (A.6) for different ranges of K by looking at the term

$$\left(\frac{\alpha(t, \tau)}{N} e^{1-\alpha(t, \tau)/N} \right)^N N^{\frac{1}{2}} = K^N N^{\frac{1}{2}}. \tag{A.8}$$

Note that K remains constant with changing N under the constraint $N/a_0 = E$. For ease of analysis we ignore the coefficient $\frac{f(t-\tau)}{E_0 \sqrt{2\pi}}$ in (A.6), which also does not change in the limit. We will show that in the limit $N \rightarrow \infty$, (A.8) is zero everywhere and infinity at a singular point for any time t .

Applying l'Hôpital's rule for $K < 1$,

$$\lim_{N \rightarrow \infty} \frac{N^{\frac{1}{2}}}{1/K^N} = \lim_{N \rightarrow \infty} \frac{\frac{1}{2} N^{-\frac{1}{2}}}{\ln(1/K)/K^N} = \lim_{N \rightarrow \infty} \frac{K^N}{2N^{1/2} \ln(1/K)} = 0, \tag{A.9}$$

and for $K \geq 1$,

$$\lim_{N \rightarrow \infty} \frac{N^{\frac{1}{2}}}{1/K^N} = \infty. \tag{A.10}$$

It remains to show that $K \leq 1$ for all τ . We would like to determine when K reaches its maximum value. As a necessary condition for an extremum we must have

$$\frac{d}{d\tau}(K) = \frac{d}{d\tau} \left(\frac{\alpha(t, \tau)}{N} \right) e^{1-\frac{\alpha(t, \tau)}{N}} \left(1 - \frac{\alpha(t, \tau)}{N} \right) = 0. \tag{A.11}$$

Since the first two terms are always strictly positive, we find that an extremum occurs at τ_{eff} , where

$$1 - \frac{\alpha(t, \tau_{\text{eff}})}{N} = 0. \tag{A.12}$$

Plugging (A.12) back into (A.7),

$$(A.13) \quad K(\tau) = 1 \cdot e^0 = 1,$$

we find $K = 1$ at the extremum. It can be easily shown that

$$(A.14) \quad \frac{d^2}{d^2\tau}(K) > 0;$$

therefore, the extremum is a maximum, and hence $K \leq 1$ for all τ . We see that in the limit as $N \rightarrow \infty$, $h(t, \tau)$ is zero everywhere for all τ except for at τ_{eff} , where $K(\tau) = 1$ and $h(t, \tau) = \infty$. Furthermore, since $\alpha(t, \tau)$ is an injective function in τ , (A.12) has a single solution; hence, τ_{eff} provides a global maximum at a given time t , which is the single nonzero solution.

Appendix B. Simulations of the dual-feedback oscillator. The oscillator is modeled by

$$(B.1) \quad \begin{aligned} \frac{1}{A(T)} \cdot \frac{dx}{dt} &= \frac{\alpha_x \left(\eta + \frac{y(t-\tau_y)}{C_y} \right)}{\left(1 + \frac{y(t-\tau_y)}{C_y} \right) \left(1 + \frac{x(t-\tau_x)}{C_x(T)} \right)^N} - \beta x(t) - \frac{\gamma_x x(t)}{R_0 + x(t) + y(t)}, \\ \frac{1}{A(T)} \cdot \frac{dy}{dt} &= \frac{\alpha_y \left(\eta + \frac{y(t-\tau_y)}{C_y} \right)}{\left(1 + \frac{y(t-\tau_y)}{C_y} \right) \left(1 + \frac{x(t-\tau_x)}{C_x(T)} \right)^N} - \beta y(t) - \frac{\gamma_y y(t)}{R_0 + x(t) + y(t)}, \end{aligned}$$

where for the temperature compensating model we have

$$(B.2) \quad C_x(T) = (C_{x,\max} - C_{x,\min}) \frac{(T/T_{\text{lac}})^b}{1 + (T/T_{\text{lac}})^b} + C_{x,\min}$$

and for the nontemperature compensating model we have

$$(B.3) \quad C_x(T) = C_x(T_O)$$

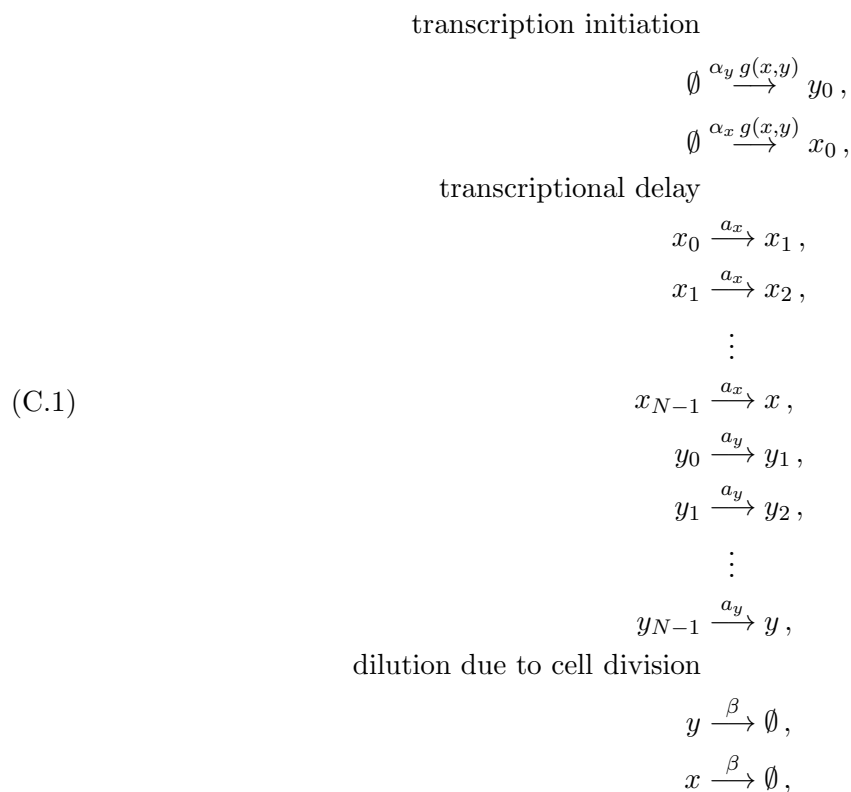
using the parameters shown in Table 1.

This system is modeled using dde23 in MATLAB with discretized delays. Using dde23, one can specify the delayed states to be used in the delay differential equation. Using the method derived in section 2, we can determine the range of delays for which we need state information. The range of delays is discretized into bins of width .05min so that state information is saved for $N \in \mathbb{Z}$ different delays, where $N = (\tau_{\max} - \tau_{\min})/.05$. The delay used in the simulation is the midpoint of each bin. At each iteration, we calculate what the time-varying delay is at that time using again the method in section 2 and find the appropriate bin. The corresponding delayed state is then fed into the delay differential equation, simulating the time-evolution of the model above with a time-varying delay.

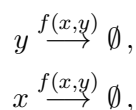
Table 1
System parameter values.

Parameter	Value
τ_x	13.5 min.
τ_y	15 min.
β	.0275 min ⁻¹
γ_x	76 (mol./cell)min ⁻¹
γ_y	76 (mol./cell)min ⁻¹
R_0	1.8 mol./cell
η	.5 (unitless)
C_y	5 mol./cell
α_x	265 (mol./cell)min ⁻¹
α_y	92.75 (mol./cell)min ⁻¹
θ	4500 K
$C_{x,\max}$	830 mol./cell
$C_{x,\min}$	50 mol./cell
T_{lac}	36°C
b	20 (unitless)
N	4

Appendix C. Gillespie simulations of the dual-feedback oscillator. We use the following reaction equations to generate Gillespie simulations:



enzymatic degradation



where

$$(C.2) \quad g(x, y) = \frac{\left(\eta + \frac{y}{C_y}\right)}{\left(1 + \frac{y}{C_y}\right) \left(1 + \frac{x}{C_x(T)}\right)^N}$$

and

$$(C.3) \quad f(x, y) = \frac{\gamma_y}{R_0 + x + y}.$$

The reaction rates are determined from the delay with equations $a_x = N/\tau_x$ and $a_y = N/\tau_y$, where we set $N = 40$ as an approximation. The remainder of the parameters are borrowed from the deterministic system. The system is simulated using the standard Gillespie algorithm [12] custom coded in MATLAB. The average period for each simulation was determined by first simulating the system for 15 periods to ensure steady state dynamics and taking the average period across the last five oscillations.

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